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Burcu F. Darst

Peggy Wan

Xin Sheng

Jeannette T. Bensen

Sue A. Ingles

See next page for additional authors

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Authors

Burcu F. Darst, Peggy Wan, Xin Sheng, Jeannette T. Bensen, Sue A. Ingles, Benjamin A. Rybicki, Barbara Nemesure, Esther M. John, Jay H. Fowke, Victoria L. Stevens, Sonja I. Berndt, Chad D. Huff, Sara S. Strom, Jong Y. Park, Wei Zheng, Elaine A. Ostrander, Patrick C. Walsh, Shiv Srivastava, John Carpten, Thomas A. Sellers, Kosj Yamoah, Adam B. Murphy, Maureen Sanderson, Dana C. Crawford, Susan M. Gapstur, William S. Bush, Melinda C. Aldrich, Olivier Cussenot, Meredith Yeager, Gyorgy Petrovics, Jennifer Cullen, Christine Neslund-Dudas, Rick A. Kittles, Jianfeng Xu, Mariana C. Stern, Zsofia Kote-Jarai, Koveela Govindasami, Anand P. Chokkalingam, Luc Multigner, Marie-Elise Parent, Florence Menegaux, Geraldine Cancel-Tassin, Adam S. Kibel, Eric A. Klein, Phyllis J. Goodman, Bettina F. Drake, Jennifer J. Hu, Peter E. Clark, Pascal Blanchet, Graham Casey, Anselm JM Hennis, Alexander Lubwama, Ian M. Thompson, Robin Leach, Susan M. Gundell, Loreall Pooler, Lucy Xia, James L. Mohler, Elizabeth TH Fontham, Gary J. Smith, Jack A. Taylor, Rosalind A. Eeles, Laurent Brureau, Stephen J. Chanock, Stephen Watya, Janet L. Stanford, Diptasri Mandal, William B. Isaacs, Kathleen Cooney, William J. Blot, David V. Conti, and Christopher A. Haiman



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European Association of Urology

Brief Correspondence

A Germline Variant at 8q24 Contributes to Familial Clustering of Prostate Cancer in Men of African Ancestry

Burcu F. Darst^a, Peggy Wan^a, Xin Sheng^a, Jeannette T. Bensen^{b,c}, Sue A. Ingles^{a,d}, Benjamin A. Rybicki^e, Barbara Nemesure^f, Esther M. John^g, Jay H. Fowke^h, Victoria L. Stevensⁱ, Sonja I. Berndt^j, Chad D. Huff^k, Sara S. Strom^k, Jong Y. Park^l, Wei Zheng^m, Elaine A. Ostranderⁿ, Patrick C. Walsh^o, Shiv Srivastava^p, John Carpten^q, Thomas A. Sellers^l, Kosj Yamoah^r, Adam B. Murphy^s, Maureen Sanderson^t, Dana C. Crawford^u, Susan M. Gapsturⁱ, William S. Bush^u, Melinda C. Aldrich^v, Olivier Cussenot^w, Meredith Yeager^j, Gyorgy Petrovics^p, Jennifer Cullen^p, Christine Neslund-Dudas^e, Rick A. Kittles^x, Jianfeng Xu^y, Mariana C. Stern^{a,d}, Zsofia Kote-Jarai^z, Koveela Govindasami^{aa}, Anand P. Chokkalingam^{bb}, Luc Multigner^{cc}, Marie-Elise Parent^{dd}, Florence Menegaux^{ee}, Geraldine Cancel-Tassin^w, Adam S. Kibel^{ff,gg}, Eric A. Klein^{hh}, Phyllis J. Goodmanⁱⁱ, Bettina F. Drake^{jj}, Jennifer J. Hu^{kk}, Peter E. Clark^{ll}, Pascal Blanchet^{cc,mm,nn}, Graham Casey^{oo}, Anselm J.M. Hennis^{ff,pp}, Alexander Lubwama^{qq}, Ian M. Thompson Jr^{rr}, Robin Leach^{rr}, Susan M. Gundell^a, Loreall Pooler^a, Lucy Xia^a, James L. Mohler^{c,ss}, Elizabeth T.H. Fontham^{tt}, Gary J. Smith^{ss}, Jack A. Taylor^{uu,vv}, Rosalind A. Eeles^{z,ww}, Laurent Brureau^{cc,mm,nn}, Stephen J. Chanock^j, Stephen Watya^{qq,xx}, Janet L. Stanford^{yy,zz}, Diptasri Mandal^{aaa}, William B. Isaacs^o, Kathleen Cooney^{bbb}, William J. Blot^m, David V. Conti^a, Christopher A. Haiman^{a,d,*}

^a Center for Genetic Epidemiology, Department of Preventive Medicine, Keck School of Medicine, University of Southern California, Los Angeles, CA, USA;

^b Department of Epidemiology, University of North Carolina at Chapel Hill, Chapel Hill, NC, USA; ^c Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, Chapel Hill, NC, USA; ^d Norris Comprehensive Cancer Center, University of Southern California, Los Angeles, CA, USA;

^e Department of Public Health Sciences, Henry Ford Hospital, Detroit, MI, USA; ^f Department of Preventive Medicine, Stony Brook University, Stony Brook, NY, USA; ^g Department of Epidemiology and Population Health and Stanford Cancer Institute, Stanford University School of Medicine, Stanford, CA, USA;

^h Division of Epidemiology, Department of Preventive Medicine, The University of Tennessee Health Science Center, Memphis, TN, USA; ⁱ Epidemiology Research Program, American Cancer Society, Atlanta, GA, USA; ^j Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institute of Health, Bethesda, MD, USA; ^k Department of Epidemiology, University of Texas M.D. Anderson Cancer Center, Houston, TX, USA; ^l Department of Cancer Epidemiology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^m Division of Epidemiology, Department of Medicine, Vanderbilt Epidemiology Center, Vanderbilt University School of Medicine, Nashville, TN, USA; ⁿ Cancer Genetics and Comparative Genomics Branch, National Human Genome Research Institute, National Institutes of Health, Bethesda, MD, USA; ^o The James Buchanan Brady Urological Institute, Johns Hopkins Hospital and Medical Institution, Baltimore, MD, USA; ^p Department of Surgery, Center for Prostate Disease Research, Uniformed Services University of the Health Sciences, Bethesda, MD, USA; ^q Department of Translational Genomics, Keck School of Medicine, University of Southern California, Los Angeles, CA, USA; ^r Department of Radiation Oncology and Cancer Epidemiology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^s Department of Urology, Northwestern University, Chicago, IL, USA; ^t Department of Family and Community Medicine, Meharry Medical College, Nashville, TN, USA; ^u Cleveland Institute for Computational Biology, Department of Population and Quantitative Health Sciences, Case Western Reserve University, Cleveland, OH, USA; ^v Department of Thoracic Surgery, Division of Epidemiology, Vanderbilt University Medical Center, Nashville, TN, USA;

^w Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^x Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^y Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^z Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{aa} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ab} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ac} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{ad} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ae} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{af} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{ag} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ah} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ai} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{aj} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ak} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{al} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{am} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{an} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ao} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{ap} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{aq} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ar} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{as} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{at} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{au} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{av} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{aw} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ax} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{ay} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{az} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ba} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{bb} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bc} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bd} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{be} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bf} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bg} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{bh} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bi} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bj} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{bk} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bl} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bm} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{bn} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bo} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bp} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{bq} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{br} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bs} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{bt} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bu} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bv} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{bw} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{bx} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{by} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{bz} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ca} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{cb} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{cc} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{cd} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ce} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{cf} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{cg} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ch} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{ci} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{cj} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{ck} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{cl} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{cm} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{cn} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

^{co} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{cp} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ^{cq} Department of Radiation Oncology, Moffitt Cancer Center and Research Institute, Tampa, FL, USA;

☆ Additional members and affiliations are presented in the Supplementary material.

* Corresponding author. Center for Genetic Epidemiology, Department of Preventive Medicine, University of Southern California, 1450 Biggy Street, Los Angeles, CA 90033, USA. Tel. +1 323 442 7755.

E-mail address: haiman@usc.edu (C.A. Haiman).

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^w CeRePP and Sorbonne Université, GRC ndeg 5, AP-HP, Tenon Hospital, Paris, France; ^x Department of Population Sciences, City of Hope Comprehensive Cancer Center, Duarte, CA, USA; ^y Program for Personalized Cancer Care and Department of Surgery, NorthShore University HealthSystem, Evanston, IL, USA; ^z The Institute of Cancer Research, Sutton, London, UK; ^{aa} Oncogenetics Team, The Institute of Cancer Research and Royal Marsden NHS Foundation Trust, Sutton, London, UK; ^{bb} School of Public Health, University of California, Berkeley, Berkeley, CA, USA; ^{cc} Inserm U1085–IRSET, Rennes, France; ^{dd} INRS-Institut Armand-Frappier, Institut National de la Recherche Scientifique, University of Quebec, Laval, Quebec, Canada; ^{ee} INSERM, Center for Research in Epidemiology and Population Health, Team Cancer-Environment, Université Paris-Saclay, Université Paris-Sud, Villejuif, France; ^{ff} Division of Urology, Brigham and Women's Hospital/Dana-Farber Cancer Institute, Boston, MA, USA; ^{gg} Washington University, St. Louis, MO, USA; ^{hh} Glickman Urological and Kidney Institute, Cleveland Clinic, Cleveland, OH, USA; ⁱⁱ SWOG Statistical Center, Fred Hutchinson Cancer Research Center, Seattle, WA, USA; ^{jj} Department of Surgery, Division of Public Health Sciences, Washington University School of Medicine, St. Louis, MO, USA; ^{kk} Sylvester Comprehensive Cancer Center and Department of Public Health Sciences, University of Miami Miller School of Medicine, Miami, FL, USA; ^{ll} Vanderbilt University Medical Center, Nashville, TN, USA; ^{mmm} University Hospital of Pointe-à-Pitre, Guadeloupe, FWI, France; ⁿⁿ French West Indies University, Pointe-à-Pitre, Guadeloupe, FWI, France; ^{oo} Center for Public Health Genomics, Department of Public Health Sciences, University of Virginia, Charlottesville, VA, USA; ^{pp} Chronic Disease Research Centre and Faculty of Medical Sciences, University of the West Indies, Bridgetown, Barbados; ^{qq} School of Public Health, Makerere University College of Health Sciences, Kampala, Uganda; ^{rr} Department of Urology, University of Texas Health Science Center at San Antonio, San Antonio, TX, USA; ^{ss} Department of Urology, Roswell Park Cancer Institute, Buffalo, NY, USA; ^{tt} School of Public Health, Louisiana State University Health Sciences Center, New Orleans, LA, USA; ^{uu} Epigenetic and Stem Cell Biology Laboratory, National Institute of Environmental Health Sciences, Research Triangle Park, NC, USA; ^{vv} Epidemiology Branch, National Institute of Environmental Health Sciences, Research Triangle Park, NC, USA; ^{www} Royal Marsden NHS Foundation Trust, London, UK; ^{xx} Uro Care, Kampala, Uganda; ^{yy} Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, WA, USA; ^{zz} Department of Epidemiology, School of Public Health, University of Washington, Seattle, WA, USA; ^{aaa} Department of Genetics, Louisiana State University Health Sciences Center, New Orleans, LA, USA; ^{bbb} Department of Medicine, Duke University of Medicine, Durham, NC, USA

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Abstract

Although men of African ancestry have a high risk of prostate cancer (PCa), no genes or mutations have been identified that contribute to familial clustering of PCa in this population. We investigated whether the African ancestry-specific PCa risk variant at 8q24, rs72725854, is enriched in men with a PCa family history in 9052 cases, 143 cases from high-risk families, and 8595 controls of African ancestry. We found the risk allele to be significantly associated with earlier age at diagnosis, more aggressive disease, and enriched in men with a PCa family history (32% of high-risk familial cases carried the variant vs 23% of cases without a family history and 12% of controls). For cases with two or more first-degree relatives with PCa who had at least one family member diagnosed at age <60 yr, the odds ratios for TA heterozygotes and TT homozygotes were 3.92 (95% confidence interval [CI] = 2.13–7.22) and 33.41 (95% CI = 10.86–102.84), respectively. Among men with a PCa family history, the absolute risk by age 60 yr reached 21% (95% CI = 17–25%) for TA heterozygotes and 38% (95% CI = 13–65%) for TT homozygotes. We estimate that in men of African ancestry, rs72725854 accounts for 32% of the total familial risk explained by all known PCa risk variants.

Patient summary: We found that rs72725854, an African ancestry-specific risk variant, is more common in men with a family history of prostate cancer and in those diagnosed with prostate cancer at younger ages. Men of African ancestry may benefit from the knowledge of their carrier status for this genetic risk variant to guide decisions about prostate cancer screening.

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Prostate cancer (PCa) is highly heritable, and having a first-degree relative with PCa is associated with a two- to three-fold increased risk [1]. With the exception of a rare *HOXB13* European-specific mutation that accounts for ~5% of hereditary PCa [2,3] and variants in DNA repair pathway genes found in ~10% of inherited PCa cases [4], genes or mutations contributing to the familial clustering of PCa remain elusive. Despite the greater risk of PCa for men of African ancestry, no single mutation has been discovered that accounts for a large fraction of the aggregation of PCa within families of African ancestry.

We and others have shown that germline variation at 8q24 is the strongest PCa risk factor across racial and ethnic populations, with multiple independent risk variants discovered in the region (127.6–129.0 Mb) [5–7]. While most associations with 8q24 variants have been observed across racial/ethnic populations, less common, ancestry-specific variants have also been detected with odds ratios

(ORs) >2.0. One such variant, rs72725854 (T risk allele frequency ~6%), is found only in men of African ancestry and is the strongest genome-wide signal for PCa (OR = 2.32; 95% confidence interval [CI] = 2.16–2.50; $p = 1.1 \times 10^{-109}$) discovered to date in this population [5]. Given the moderate effect size conveyed by this variant, we investigated whether the T allele is associated with PCa family history and age at diagnosis, characteristics that indicate a strong genetic influence of disease onset.

This study included 9052 PCa cases unselected for PCa family history (median age = 64 [interquartile range = 12] yr, 20% with a PCa family history), of which 2041 had high-risk disease and 692 lethal disease, and 8595 controls (median age = 64 [interquartile range = 13] yr, 9.0% with a PCa family history; Supplementary Tables 1 and 2). Among cases, 23.7% carried at least one copy of the T allele versus 11.6% of controls; the ORs were 2.29 (95% CI = 2.10–2.49) for TA heterozygotes and 5.04 (95% CI = 3.36–7.55) for TT homozygotes (Table 1 and

Table 1 – Prostate cancer risk associated with the germline variant rs72725854 by family history^{a,b} of prostate cancer and age at diagnosis.

| Group | N | Risk allele frequency (%) | Carrier frequency (%) | Homozygote genotype frequency (%) | Heterozygote genotype | | | Homozygote variant genotype | | |
|---|------|---------------------------|-----------------------|-----------------------------------|-----------------------|------------------|---------|-----------------------------|----------------------|---------|
| | | | | | Frequency (%) | OR (95% CI) | p value | Frequency (%) | OR (95% CI) | p value |
| Controls (reference group) | 8595 | 6.0 | 11.6 | 88.3 | 11.3 | | | 0.35 | | |
| Cases | 9052 | 12.6 | 23.7 | 76.3 | 22.3 | 2.29 (2.10–2.49) | 6.7E-81 | 1.4 | 5.04 (3.36–7.55) | 5.2E-15 |
| Cases with at least 1 first-degree relative with prostate cancer | 1795 | 14.8 | 27.4 | 72.6 | 25.1 | 2.67 (2.34–3.04) | 1.6E-48 | 2.3 | 8.66 (5.22–14.37) | 6.8E-17 |
| Cases with no family history of prostate cancer | 6125 | 11.9 | 22.7 | 77.4 | 21.5 | 2.17 (1.97–2.38) | 1.1E-59 | 1.2 | 4.31 (2.77–6.71) | 8.8E-11 |
| Cases with age at diagnosis <60 yr | 2923 | 15.1 | 28.2 | 71.8 | 26.2 | 2.63 (2.32–2.97) | 5.9E-52 | 2.0 | 6.84 (4.09–11.44) | 2.4E-13 |
| Cases with age at diagnosis ≥60 yr | 6129 | 11.4 | 21.6 | 78.4 | 20.4 | 2.10 (1.90–2.31) | 2.6E-49 | 1.2 | 4.22 (2.69–6.63) | 4.2E-10 |
| Cases with at least 1 first-degree relative with prostate cancer and case age at diagnosis <60 yr | 688 | 16.9 | 30.8 | 69.2 | 27.9 | 2.85 (2.33–3.49) | 1.4E-24 | 2.9 | 11.56 (6.00–22.27) | 2.5E-13 |
| Familial cases | 143 | 17.8 | 32.2 | 67.8 | 28.7 | 3.33 (2.29–4.85) | 3.3E-10 | 3.5 | 13.92 (5.15–37.63) | 2.1E-07 |
| Familial cases with ≥2 first-degree relatives with prostate cancer | 76 | 19.1 | 32.9 | 67.1 | 27.6 | 3.23 (1.92–5.41) | 8.8E-06 | 5.3 | 21.20 (7.04–63.83) | 5.6E-08 |
| Familial cases with ≥2 first-degree relatives with prostate cancer and age at diagnosis <60 yr (any relative) | 51 | 23.5 | 39.2 | 60.8 | 31.4 | 3.92 (2.13–7.22) | 1.2E-05 | 7.8 | 33.41 (10.86–102.84) | 9.5E-10 |

CI = confidence interval; OR = odds ratio.
^a Controls are limited to 8594 for comparisons with the 143 familial cases, as one control is a full sibling of one of the familial cases.
^b Family history in nonfamilial cases and controls includes first- or second-degree relatives with prostate cancer.

Supplementary Fig. 1). The percentage of cases carrying the T allele was significantly greater for men with a PCa family history (27.4% in those with a first/second-degree relative with PCa vs 22.7% in those without, $p = 0.002$) and for men diagnosed at age <60 yr (28.2% vs 21.6% for those aged ≥60 yr at diagnosis, $p = 0.002$; Table 1). The mean age at diagnosis for TT homozygotes was 61.1 yr (standard deviation [SD]=8.7) versus 62.7 yr (SD=9.1) for TA heterozygotes and 64.3 yr (SD=8.9) for AA homozygotes ($p = 5.7E-14$).

The percentage of cases carrying the T allele was highest among men with both a family history and an early diagnosis (30.8%; Table 1 and Supplementary Fig. 1). For men with a PCa family history and aged <60 yr at diagnosis versus controls, the ORs were 2.85 (95% CI=2.33–3.49) for TA heterozygotes and 11.56 (95% CI=6.00–22.27) for TT homozygotes.

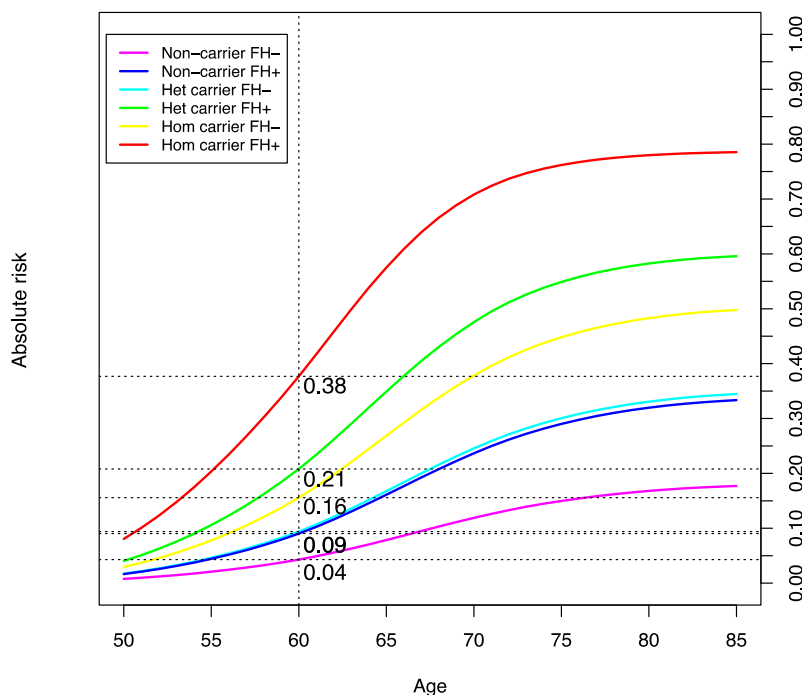
The T allele was over-represented in advanced PCa cases (Supplementary Table 3), with the percentage of men carrying the T allele ranging from 26.0% for lethal PCa (metastatic disease, prostate-specific antigen [PSA] > 100 ng/mL, or PCa death), 25.4% for high-risk disease (stage T3/T4, Gleason 8–10, or PSA = 20–100 ng/mL), 24.6% for intermediate-risk disease (Gleason 7, stage T1/T2, and PSA = 10–20 ng/mL), and 21.4% for low-risk disease (Gleason < 7, stage T1/T2, and PSA < 10 ng/mL; heterogeneity $p = 0.03$). This pattern was consistent in cases with (35.1%, 28.8%, 27.4%, and 27.2%, respectively; heterogeneity $p = 0.2$) and without (24.8%, 24.2%, 24.1%, and 19.4%, respectively; heterogeneity $p = 0.01$) a PCa family history.

Given the enrichment of the T allele in cases with a PCa family history and early-onset disease, we examined a dose-response relationship between the risk allele and strength of

family history using an independent sample of 143 high-risk families (Supplementary material and Supplementary Tables 1 and 2). Among affected probands, 32.2% carried the risk allele, with 3.5% being homozygotes. Comparing these familial cases with the 8595 controls, the ORs for TA heterozygotes and TT homozygotes were 3.33 (95% CI=2.29–4.85) and 13.92 (95% CI=5.15–37.63), respectively (Table 1). The percentage of cases carrying the T allele was greater for those with two or more first-degree relatives with PCa ($n = 76$, 32.9%) and for those who also had at least one family member aged <60 yr at diagnosis ($n = 51$, 39.2%). For this latter subset of cases, the ORs for TA heterozygotes and TT homozygotes were 3.92 (95% CI=2.13–7.22) and 33.41 (95% CI=10.86–102.84), respectively.

We estimated the absolute risk of PCa based on the log-additive effects of rs72725854 and family history (Supplementary material and Supplementary Table 4). Among men without a family history, the absolute risks for PCa by age 60 yr for TA heterozygotes and TT homozygotes were 9.0% (95% CI=8.6–10%) and 16% (95% CI=8.7–23%), respectively, versus 4.3% (95% CI=4.1–4.5%) for nonrisk allele carriers (Fig. 1 and Supplementary Table 5). Among men with a PCa family history, the absolute risk by age 60 yr reached 21% (95% CI=17–25%) for TA heterozygotes and 38% (95% CI=13–65%) for TT homozygotes, versus 9.0% (95% CI=8.2–9.8%) for nonrisk allele carriers. Lifetime absolute risk of PCa for heterozygotes and homozygotes with a family history reached 60% (95% CI=53–66%) and 79% (95% CI=57–100%), respectively.

While the T allele is more common in men with a higher percentage of global African ancestry (Supplementary Fig. 2 and Supplementary material), its frequency varies widely



| Group | #Controls | #Cases | OR (95% CI) | p value |
|-----------------------------|-----------|--------|--------------------|---------|
| Noncarrier FH- (ref. group) | 5875 | 4747 | | |
| Noncarrier FH+ | 675 | 1304 | 2.15 (1.93–2.41) | 6.8E-42 |
| Heterozygote carrier FH- | 728 | 1316 | 2.25 (2.02–2.51) | 2.9E-48 |
| Heterozygote carrier FH+ | 96 | 450 | 5.31 (4.19–6.73) | 2.5E-43 |
| Homozygote carrier FH- | 22 | 72 | 3.85 (2.29–6.50) | 4.1E-07 |
| Homozygote carrier FH+ | 5 | 41 | 10.77 (4.15–27.95) | 1.1E-06 |

Fig. 1 – Absolute risk of prostate cancer by rs72725854 genotype and family history. Absolute risks are estimated using the odds ratios for each genotype/family history category, which are indicated in the table, combined with mortality and incidence rates for African-American men (see the Supplementary material). CI = confidence interval; FH+ = participants with at least one first- or second-degree relative with prostate cancer; FH- = participants with no family history of prostate cancer; Hom = homozygote; Het = heterozygote; OR = odds ratio.

between African populations. Based on genotype data from the current and previously published studies and from the 1000 Genomes Project, the T allele is found to range from 2.5% in Acholi men from North-Central Uganda to 15.0% in men from the Democratic Republic of the Congo (Supplementary Table 6), suggesting that in some populations, over 25% of men may be carriers of the risk allele (compared with 11.6% of unaffected men in our study, the majority of whom were African American).

We also evaluated four other 8q24 risk variants found to be associated with PCa in men of African ancestry [5] and found that these variants were not over-represented in men with a PCa family history (Supplementary Table 7). When combining the five 8q24 variants in a genetic risk score (GRS; Supplementary material), the PCa OR for the top 10% versus median 40–60% GRS was 2.33 (95% CI = 2.08–2.62). This OR was considerably diminished after excluding rs72725854 to 1.54 (95% CI = 1.35–1.77; Supplementary Fig. 3). This association between PCa and a GRS excluding rs72725854 was similar between those with and without a PCa family history and those diagnosed at younger and older ages (Supplementary Table 8). Findings were similar for a GRS of the 180 known PCa risk variants, with ORs for the top 10% versus median 40–50% GRS of 2.84

(95% CI = 2.53–3.18) including and 2.28 (95% CI = 2.03–2.56) excluding rs72725854 (Supplementary Fig. 3 and Supplementary Table 8). These results suggest that rs72725854 is uniquely over-represented in men with PCa who have a family history and are diagnosed at a younger age.

We estimate that in men of African ancestry, the 180 known genetic risk variants collectively explain 29–38% of the familial relative risk of PCa (based on observed familial risk to first-degree relatives of PCa cases, ranging from 2.0 to 2.5, as described in the Supplementary material and Supplementary Table 9). The five African ancestry 8q24 variants explain 14–18% of familial risk of PCa (accounting for 49% of the total familial risk explained by the 180 variants; Supplementary Table 9), which is twice the amount of familial risk explained by 8q24 in men of European ancestry (~25%) [8]. [1–11] The variant rs72725854 alone explains 9.2–12% of the familial relative risk (32% of the total familial risk explained by the 180 variants). This is a considerably larger contribution to familial risk than any other known risk variant (Supplementary Table 10), including the rare *HOXB13* PCa risk variant found only in men of European ancestry, which is estimated to explain ~5% of the total familial risk explained by all known PCa risk variants [8] and accounts for ~5% of all familial cases [3]. Our findings support the importance of

8q24 germline variation, while highlighting the much larger contribution of 8q24 risk variants to familial risk for men of African versus European ancestry.

We demonstrate that the germline variant rs72725854 is significantly enriched among PCa cases of African ancestry with a PCa family history. The T allele was observed in 32% of familial cases, with a 33-fold increase in the percentage of high-risk familial cases who were homozygous risk allele carriers (7.8%) compared with controls (0.35%). Although the biological and functional relevance of this risk variant has not been determined, it is located in a noncoding intergenic region near Prostate Cancer Associated Transcript 1 (*PCAT1*) and other PCa-associated long noncoding RNAs implicated in the promotion of PCa through regulation of the downstream *MYC* oncogene [9,10]. The lack of this variant in European ancestry populations emphasizes differences underlying the genetic architecture of PCa in men of African versus European descent.

Given the association of the germline variant rs72725854 with a high absolute PCa risk and an increased risk of aggressive and lethal disease, men of African ancestry may benefit from the knowledge of their carrier status for this genetic risk variant to guide decisions about PCa screening. Carriers of the rare *HOXB13* mutation or *BRCA2* mutations have been recommended to start PCa screening with a baseline PSA measurement at age 40 yr or 10 yr prior to the diagnosis of the youngest PCa patient in the family [11]. Men of African ancestry carrying the rs72725854 risk allele could benefit from similar screening recommendations. Further research is needed, particularly a prospective investigation of rs72725854 T allele carriers, to determine when and how to incorporate rs72725854 into PCa screening and whether combining rs72725854 genotype information with screening measures, such as PSA, the 4Kscore, the Stockholm-3 model, or the Prostate Health Index, could effectively identify PCa and aggressive disease earlier in men of African ancestry.

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Study concept and design: Haiman.

Acquisition of data: Wan, Sheng, Xia, Gundell, Pooler, Bensen, Ingles, Rybicki, Nemesure, John, Fowke, Stevens, Berndt, Huff, Strom, Park, Zheng, Ostrander, Walsh, Srivastava, Carpten, Sellers, Yamoah, Murphy, Sanderson, Crawford, Gapstur, Bush, Aldrich, Cussenot, Petrovics, Cullen, Neslund-Dudas, Kittles, Xu, Stern, Kote-Jarai, Govindasami, Chokkalingam, Multigner, Parent, Menegaux, Cancel-Tassin, Kibel, Klein, Goodman, Drake, Hu, Clark, Blanchet, Casey, Hennis, Lubwama, Thompson Jr, Leach, Gundell, Pooler, Mohler, Fontham, Smith, Taylor, Eeles, Brureau, Chanock, Watya, Stanford, Mandal, Isaacs, Cooney, Blot, Conti, Haiman.

Analysis and interpretation of data: Darst, Conti, Haiman, Wan, Sheng, Xia, Yeager.

Drafting of the manuscript: Darst, Conti, Haiman.

Critical revision of the manuscript for important intellectual content: Sheng, Sellers, John, Chanock, Crawford.

Statistical analysis: Darst, Conti, Wan, Sheng, Xia.

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Appendix A. Supplementary data

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References

- [1] Stanford JL, Ostrander EA. Familial prostate cancer. *Epidemiol Rev* 2001;23:19–23.
- [2] Ewing CM, Ray AM, Lange EM, et al. Germline mutations in *HOXB13* and prostate-cancer risk. *N Engl J Med* 2012;366:141–9.
- [3] Xu J, Lange EM, Lu L, et al. *HOXB13* is a susceptibility gene for prostate cancer: results from the International Consortium for Prostate Cancer Genetics (ICPCG). *Hum Genet* 2013;132:5–14.
- [4] Giri VN, Hegarty SE, Hyatt C, et al. Germline genetic testing for inherited prostate cancer in practice: implications for genetic testing, precision therapy, and cascade testing. *Prostate* 2019;79:333–9.
- [5] Conti DV, Wang K, Sheng X, et al. Two novel susceptibility loci for prostate cancer in men of African ancestry. *J Natl Cancer Inst* 2017;109:djx084.
- [6] Freedman ML, Haiman CA, Patterson N, et al. Admixture mapping identifies 8q24 as a prostate cancer risk locus in African-American men. *Proc Natl Acad Sci U S A* 2006;103:14068–73.
- [7] Haiman CA, Patterson N, Freedman ML, et al. Multiple regions within 8q24 independently affect risk for prostate cancer. *Nat Genet* 2007;39:638–44.
- [8] Matejic M, Saunders EJ, Dadaev T, et al. Germline variation at 8q24 and prostate cancer risk in men of European ancestry. *Nat Commun* 2018;9:4616.
- [9] Prensner JR, Chen W, Han S, et al. The long non-coding RNA *PCAT-1* promotes prostate cancer cell proliferation through cMyc. *Neoplasia* 2014;16:900–8.
- [10] Kim T, Cui R, Jeon YJ, et al. Long-range interaction and correlation between *MYC* enhancer and oncogenic long noncoding RNA *CARLO-5*. *Proc Natl Acad Sci U S A* 2014;111:4173–8.
- [11] Giri VN, Knudsen KE, Kelly WK, et al. Role of genetic testing for inherited prostate cancer risk: Philadelphia Prostate Cancer Consensus Conference 2017. *J Clin Oncol* 2018;36:414–24.