Secondary Release of Exosomes From Astrocytes Contributes to the Increase in Neural Plasticity and Improvement of Functional Recovery After Stroke in Rats Treated With Exosomes Harvested From MicroRNA 133b-Overexpressing Multipotent Mesenchymal Stromal Cells

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Secondary Release of Exosomes From Astrocytes Contributes to the Increase in Neural Plasticity and Improvement of Functional Recovery After Stroke in Rats Treated With Exosomes Harvested From MicroRNA 133b-Overexpressing Multipotent Mesenchymal Stromal Cells

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We previously demonstrated that multipotent mesenchymal stromal cells (MSCs) that overexpress microRNA 133b (miR-133b) significantly improve functional recovery in rats subjected to middle cerebral artery occlusion (MCAO) compared with naive MSCs and that exosomes generated from naive MSCs mediate the therapeutic benefits of MSC therapy for stroke. Here we investigated whether exosomes isolated from miR-133b-overexpressing MSCs (Ex-miR-133b+) exert amplified therapeutic effects. Rats subjected to 2 h of MCAO were intra-arterially injected with Ex-miR-133b+, exosomes from MSCs infected by blank vector (Ex-Con), or phosphate-buffered saline (PBS) and were sacrificed 28 days after MCAO. Compared with the PBS treatment, both exosome treatment groups exhibited significant improvement of functional recovery. Ex-miR-133b+ treatment significantly increased functional improvement and neurite remodeling/brain plasticity in the ischemic boundary area compared with the Ex-Con treatment. Treatment with Ex-miR-133b+ also significantly increased brain exosome content compared with Ex-Con treatment. To elucidate mechanisms underlying the enhanced therapeutic effects of Ex-miR-133b+, astrocytes cultured under oxygen- and glucose-deprived (OGD) conditions were incubated with exosomes harvested from naive MSCs (Ex-Naive), miR-133b downregulated MSCs (Ex-miR-133b−), and Ex-miR-133b+. Compared with the Ex-Naive treatment, Ex-miR-133b+ significantly increased exosomes released by OGD astrocytes, whereas Ex-miR-133b− significantly decreased the release. Also, exosomes harvested from OGD astrocytes treated with Ex-miR-133b+ significantly increased neurite branching and elongation of cultured cortical embryonic rat neurons compared with the exosomes from OGD astrocytes subjected to Ex-Con. Our data suggest that exosomes harvested from miR-133b-overexpressing MSCs improve neural plasticity and functional recovery after stroke with a contribution from a stimulated secondary release of neurite-promoting exosomes from astrocytes.

Key words: MicroRNA 133b (miR-133b); Exosome; Multipotent mesenchymal stromal cell (MSCs); Neural plasticity; Astrocyte; Stroke; Functional recovery

INTRODUCTION

Multipotent mesenchymal stromal cells (MSCs) enhance functional recovery in experimental stroke models by amplifying neurogenesis, angiogenesis, and neural plasticity.1–5 Our prior in vitro and in vivo studies indicated that the therapeutic benefits of MSC-based therapies are mediated by exosome-enriched extracellular particles that transfer microRNAs (miRNAs) from MSCs to neural cells.6–10 This exosome transfer of miRNAs evokes neurite remodeling and brain plasticity and subsequently benefits functional recovery.6–8. Treatment of rats subjected to middle cerebral artery occlusion (MCAO) with MSCs over-expressing microRNA 133b (miR-133b) augments neurite remodeling and brain plasticity and subsequently improves functional recovery.6. Whether treatment of stroke with cell-free exosomes enriched with miR-133b affects brain plasticity and neurological recovery has not been investigated. Exosomal release is regulated by specific physiological and pathological conditions and by multivesicular body (MVB) fusion with the plasma membrane. Factors...
that impact MVB fusion affect exosome secretion, for example, intracellular calcium14–16, K+-induced depolarization15, and glutaminergic activity17. Extracellular exosomes may also impact the exosomes released by the recipient cells. An in vitro study has shown that exosomes from normal human mammary epithelial cells inhibit exosome secretion from breast cancer cells by means of negative feedback control after intake of the epithelial cell exosomes by breast cancer cells18.

Here we intra-arterially (IA) administered miR-133b-enriched exosomes derived from MSCs to rats subjected to MCAO and investigated whether these exosomes exert amplified therapeutic effects compared with the control MSC exosomes. We demonstrated that exosomes tailored with enriched miR-133b from MSCs enhance neurological recovery and plasticity poststroke compared with the control MSC exosomes. We further demonstrated, for the first time, using in vivo and in vitro approaches, that miR-133b-enriched exosomes amplify neurite outgrowth poststroke by stimulating secondary release of astrocytic exosomes.

**MATERIALS AND METHODS**

All experimental procedures were carried out in accordance with the National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Animal Care and Use Committee (IACUC) of the Henry Ford Health System. All exosome-enriched extracellular vesicles (EVs) were collected using multistep ultracentrifugation, and their sizes were verified by transmission electron microscopy as well as Western blot detection of common exosomal proteins, CD6319,20 and CD8121,22. For simplification, in the present study, we refer to these EVs as exosomes.

**Production for miR-133b Knockin or Knockdown MSC Exosomes**

MSCs (three to four passages (P3–4)) were dissociated from Wistar rat bone marrow, cultured, and then infected with lentivirus, as previously described6,23–25. The lentivirus was constructed with the vectors of LentimiRa-GFP-hsa-miR-133b Vector (mh10170; pre-miR-133b was inserted for miR-133b knockin, Lenti-miR-133b+; Applied Biological Materials Inc., Richmond, Canada), pLenti-III-miR-GFP Control Vector (m001; vector for miR-133b knockin control, Lenti-miR-CON+; Applied Biological Materials Inc.), miRZip-133b anti-miR-133b miRNA construct (MZIPI33b-PA-1; miR-133b inhibitor inserted for miR-133b knockdown, Lenti-miR-133b−; System Biosciences, LLC, Palo Alto, CA, USA), and pGreenPuro Scramble Hairpin Control Construct (MZIPO000-PA-1; vector for miR-133b knockdown control, Lenti-miR-CON−; System Biosciences, Inc.), respectively, according to the manufacturer’s suggested protocol. MSCs were infected with lentivirus when they achieved 80%–90% confluence and were selected by puromycin for constant gene expression. Correspondingly, the MSCs generated are referred to as miR-133b+MSCs, miR-CON+MSCs, miR-133b−MSCs, and miR-CON−MSCs, respectively. For the harvesting of exosomes, we replaced conventional culture medium with an exosome-depleted fetal bovine serum (FBS; EXO-FBS-250A-1; System Biosciences, Inc.) medium when the cells reached 60%–80% confluence. After an additional 24 h of culturing, the media were then collected and centrifuged at 3,000×g for 30 min to remove the dead cells and large cell apoptotic bodies; then the upper media were stored in a −80°C freezer for exosome isolation.

**MCAO Model, Exosome Treatment, and Behavioral Tests**

In our previous in vivo stroke studies, we administered exosomes intravenously7. In addition, we have shown that direct application of MSC exosomes on cultured cortical neurons induces neurite outgrowth via transfer of miR-133b8. In the current study, in order to reduce the potential liver and lung sequestration of exosomes26,27 and to optimize the MSC exosome localization with neural parenchymal cells, we administered the exosomes using an IA approach.

Adult male Wistar rats (weighing 270–300 g) were purchased from Charles River Laboratories (Wilmington, MA, USA) and were subjected to 2 h of MCAO using a method of intraluminal vascular occlusion as modified in our laboratory28,29. Briefly, rats were anesthetized with 3.5% isoflurane for induction and maintained by spontaneously respired 1.0% isoflurane in a 2:1 N2O/O2 mixture using a face mask. The rectal temperature was kept at 37 ± 1°C throughout the surgical procedure using a feedback-regulated water-heating system. MCAO was induced by advancing a 4-0 surgical nylon suture, with its tip rounded by heating near a flame, from the external carotid artery into the lumen of the internal carotid artery (ICA) to block the origin of the right middle cerebral artery (MCA). After 2 h of MCAO, animals were reanesthetized with isoflurane, and reperfusion was performed by withdrawal of the suture until the tip cleared the lumen of the external carotid artery. The procedure provides a focal infarct in the unilateral striatum and cortex28,30.

Twenty-four hours after induction of stroke, rats were randomly administered exosomes (3×1011 particles, comparable to 100 µg of total exosome protein, per rat) harvested from miR-CON+MSCs (Ex-Con) or from miR-133b−MSCs (Ex-miR-133b−) dissolved into 0.5 ml of phosphate-buffered saline (PBS; Thermo Fisher Scientific Inc., Grand Island, NY, USA) or 0.5 ml of PBS as vehicle control (n=6/group).

For evaluation of functional recovery, a foot fault test31 and a modified neurologic severity score (mNSS) test32 were administered to the rats at 1, 3, 7, 14, and 28 days poststroke. For all behavioral tests, behavioral data were collected before surgery, at 1, 3, 7, 14, and 28 days poststroke.
were performed prior to MCAO and 1, 3, 7, 14, 21, and 28 days after MCAO by an investigator blinded to the treatments. All rats were sacrificed 28 days after MCAO. Rat brains were snap frozen in liquid nitrogen, and the frozen coronal sections were cut (40-µm thickness for molecular studies, e.g., Western blot and exosome extraction; 8-µm thickness for immunohistochemical staining) and stored in −80°C for subsequent analysis.

**Exosome Isolation and Quantification**

Exosome isolation from the cell cultured media was performed at 4°C via multistep centrifugation, as previously described. Briefly, the stocked cultured cell media were thawed and followed by vortex mixing for 1 min before centrifugation, and then centrifuged at 10,000 × g for 30 min to remove large debris. The supernatants were filtered through a 0.22-µm filter (SLMP025SS; EMD Millpore Corporation, Billerica, MA, USA) to remove small cell debris, and the resulting media were further centrifuged at 100,000 × g for 2 h. By this step, the pellets primarily contained exosomes. The final exosome pellets harvested from 1 × 10^7 cells were identified, as previously described, and resuspended in 30 to 50 µl of PBS and stored at −80°C for further use.

Exosomes in the ischemic boundary zone (IBZ) were extracted by following a previously described protocol with modification. Briefly, the IBZ tissues from frozen rat brain sections were collected and digested with 20 U/ml papain (LS 03126; Worthington Biochemical Corp., Lakewood, NJ, USA) in Hibernate E solution (20 µl/mg brain tissue; Life Technologies Corporation, Grand Island, NY, USA) for 15 min at 37°C. The brain tissue was gently homogenized in two volumes of cold Hibernate E solution. The brain homogenate was sequentially filtered through a Falcon™ 40-µm mesh filter (Catalog No. 08-771-1; Thermo Fisher Scientific, Hampton, NH, USA). Exosomes were isolated from the filtrate, as previously described. Briefly, the filtrate was sequentially centrifuged at 300 × g for 10 min at 4°C, 2,000 × g for 10 min at 4°C, 10,000 × g for 30 min at 4°C and filtered through 0.2-µm syringe filters to discard cells, membranes, and debris. The supernatant was centrifuged at 100,000 × g × 30 min at 4°C to separate oligodendrocytes from the astrocytes. Medium was changed and flask was placed on a shaker platform at a horizontal position with the medium covering the cells and were shaken at 350 rpm for 6 h at 35°C to separate oligodendrocytes from the astrocytes. Medium was changed and flask was placed on the shaker for an additional 18 h. Afterward, medium was again changed and shook for an additional 24 h. When confluent, cells were passed and used for MSC exosome treatment. To mimic the in vivo ischemic condition, an in vitro oxygen- and glucose-deprived (OGD) culture system on astrocytes was employed, as previously described. Primary cultured astrocytes were washed twice with PBS before MSC exosome treatment, and then the media containing exosomes (3 × 10^10 particles/ml) or media alone were added to the cultured cells.

**Neurite Outgrowth Assay**

Primary cultures of cortical neurons were harvested from embryonic day 18 Wistar rats, as previously described. Pregnant rats purchased from Charles River Laboratories were euthanized under deep pentobarbital anesthesia. Embryos were removed, and the cerebral cortex was dissected out, washed in Hank’s balanced salt solution (HBSS; Gibco, Grand Island, NY, USA), and dissociated by a combination of 0.125% trypsin digestion for 15 min, then mechanically triturated 20 times. The triturated cells were passed through a 40-µm cell strainer and counted to obtain a concentration of 3 × 10^7 cells/ml.

For the neurite outgrowth assay, the neurons were then plated overnight onto a poly(l-lysine)-coated 24-well plate at a density of 5,000 cells per well in DMEM containing 5% FBS and antibiotics. After 24 h, cells were washed three times with PBS and treated with various astrocytic-derived exosomes dissolved at a concentration of 3 × 10^10 particles/ml into the serum-free neurobasal medium (Gibco) supplemented with 2% B-27 (Gibco), 500 µM L-glutamine, and antibiotics (Catalog No. G6784; Sigma-Aldrich, St. Louis, MO, USA). After 3 days, neurons were washed and fixed with 4% paraformaldehyde (PFA) and immunofluorescence staining for β-III-tubulin (TuJ1) for neurite outgrowth analysis, as previously described. Briefly, to analyze neurite outgrowth, TuJ1+ cells were digitized using a 20× objective (Carl Zeiss Microscopy, LLC, Thornwood, NY, USA) via the Micro

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**In Vitro Primary Astrocyte Culture and Treatments**

Extracts from cortices of embryonic day 18 Wistar rats (either sex; Charles River Laboratories) were dissociated into a cell suspension using mechanical digestion. After digestion, cells were plated in 75-cm² tissue culture flasks at a concentration of 1.5 × 10^6 cells in Dulbecco’s modified Eagle’s medium (DMEM; Catalog No. 11995-065; Thermo Fisher Scientific) containing 20% FBS. The flasks were incubated at 37°C in a moist 5% CO₂, 95% air atmosphere for 48–72 h before moving. The medium was changed every 48–72 h. After incubating the primary cultures for 7–9 days, the flasks were placed on a shaker platform in a horizontal position with the medium covering the cells and were shaken at 350 rpm for 6 h at 35°C to separate oligodendrocytes from the astrocytes. Medium was changed and flasks were placed on the shaker for an additional 18 h. Afterward, medium was again changed and shook for an additional 24 h. When confluent, cells were passed and used for MSC exosome treatment. To mimic the in vivo ischemic condition, an in vitro oxygen- and glucose-deprived (OGD) culture system on astrocytes was employed, as previously described. Primary cultured astrocytes were washed twice with PBS before MSC exosome treatment, and then the media containing exosomes (3 × 10^10 particles/ml) or media alone were added to the cultured cells.
Computer Imaging Device (MCID) analysis system (Imaging Research, St. Catharines, Canada). Neurite outgrowth was quantified using a software program developed in our laboratory that includes measurements of the number and length of branches\(^40\). At least 50 TuJ\(^1\) cells, distributed in nine random fields per well, and triple wells per group were measured, and three independent experiments were performed. All measurements were performed by experimenters blinded to each culture condition.

**miRNA Assay**

For the measurement of miR-133b in exosomes from cultured cells, samples were lysed in QIAzol reagents, and the total RNA was isolated using the miRNeasy Mini Kit (Qiagen, Valencia, CA, USA). By real-time polymerase chain reaction (RT-PCR), we detected the miR-133b level. Briefly, miRNAs were reverse transcribed with the miRNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA, USA), and PCR amplification was performed with the TaqMan miRNA Assay Kit (which is specific for mature miRNA sequences; Applied Biosystems) according to the manufacturer’s protocols, with U6 snRNA as an internal control.

**Histopathology and Immunohistochemistry**

To determine neurite remodeling in the IBZ, adjacent frozen coronal sections of rat brains were used for staining the following markers, and positive staining within the IBZ was measured under light microscopy. Bielschowsky silver (silver nitrate; stains neuronal processes\(^41\); Sigma-Aldrich) combined with Luxol fast blue (stains myelin sheath\(^42\); Sigma-Aldrich) histochemistry staining as well as immunostaining with antibodies against the phosphorylated epitope of neurofilament heavy polypeptide (NF-H), clone SMI 31 (SMI 31; reacts broadly with thick and thin axons and some dendrites\(^43\)), and synaptophysin (a marker for synapses, since synaptophysin is ubiquitously present at the synapses\(^44\)) were employed, respectively. Briefly, for immunostaining, adjacent frozen brain sections were incubated with the primary antibodies against SMI 31 (1:500; ab82259; Abcam, Cambridge, MA, USA), synaptophysin (1:100; MAB5258; Chemicon, Billerica, MA, USA), and BrdU (1:100; Roche Diagnostics Corporation, Indianapolis, IN, USA), followed with corresponding horseradish peroxidase (HRP)-conjugated secondary antibodies (Abcam) and 3,3′-diaminobenzidine (DAB; Sigma-Aldrich) developing, respectively. Positive staining within nine areas (four from the cortex, four from the striatum, and one from the corpus callosum) selected along the IBZ in these groups were digitized under a 40× objective (BX40; Olympus Optical, Waltham, MA, USA) using a 3-CCD color video camera (DXC-970MD; Sony, Teaneck, NJ, USA) interfaced with the MCID™ software\(^45\). For the analysis of neurite remodeling, based on evaluation of an average of three histology slides (8-μm thick, interval of every 10th slide) from the standard block of each animal, the percentage of positive staining area to the whole area of each image taken within the IBZ was analyzed using the MCID™ software.

To detect astrocytic expression of Rab9 effector protein with kelch motifs (RABEPK) in the IBZ, double immunofluorescence staining was employed to identify the cellular colocalization of RABEPK (1:100; sc-107078; Santa Cruz Biotechnology, Dallas, TX, USA) with glial fibrillary acidic protein (GFAP; a marker of reactive astrocyte; 1:10,000; Z0334; Dako, Carpinteria, CA, USA), followed by their corresponding second antibody staining [fluorescein isothiocyanate (FITC) labeled for RABEPK and Cy3 labeled for GFAP].

**Statistical Analysis**

Data are summarized and presented using mean± standard error (SE). Post hoc tests were used for subgroup analysis following analysis of variance (ANOVA). The Global test using the generalized estimating equation (GEE) was employed to evaluate the MSC exosome treatment effects influenced by miR-133b enrichment on functional recovery\(^46,47\). Repeated-measures analysis was used to evaluate data with repeated measurements over time (e.g., functional tests) or histological evaluation on multiple regions per subject. Analysis began testing for the factor interaction, followed by testing the main factor effect if no interaction was detected at the 0.05 level, and/or a subgroup analysis was performed if the interaction or main effect was detected at the 0.05 level\(^48\). All statistical analyses were conducted using the SAS software (version 9.2; SAS Institute, Cary, NC, USA).

**RESULTS**

**miR-133b Level Modified by Lentivirus Introduction of Pre-miR-133b and miR-133b Inhibitors**

Quantitative RT-PCR analysis showed that mature miR-133b levels in the MSCs infected by lentivirus carrying pre-miR-133b were increased and, concomitantly, miR-133b levels in exosomes harvested from these MSCs significantly increased (Table 1). Correspondingly, the miR-133b levels in MSCs and harvested exosomes were significantly decreased by introducing an miR-133b inhibitor into MSCs (Table 1). There was no statistical difference in miR-133b levels among naive MSCs and MSCs infected by blank vector controls (miR-CON+MSCs and miR-CON−MSCs) as well as in their corresponding exosomes (Table 1).

**miR-133b-Enriched Exosomes Improve Neurological Outcome After MCAO**

Tailored exosomes enriched with miR-133b as well as the exosomes harvested from blank vector control
miR-133b INCREASES BRAIN SECONDARY EXOSOME RELEASE

MSCs administered IA to rats subjected to 2 h of transient MCAO significantly reduced neurological deficits compared with the PBS-treated rats. Foot fault and mNSS tests were performed to assess the sensorimotor function. All rats exhibited severe sensorimotor deficit at postoperative day 1, followed by gradual improvement within the 4-week experimental course. Compared with the PBS treatment, both exosome treatment groups exhibited significant improvement of functional recovery in the foot fault test (Fig. 1A) and mNSS (Fig. 1B); however, significant improvement was detected at 7 and 14 days after MCAO in the Ex-miR-133b+ and the Ex-Con treatment groups, respectively. Compared with the Ex-Con, the Ex-miR-133b+ treatment significantly increased functional improvement (p<0.05 after day 21, respectively) (Fig. 1A and B).

miR-133b-Enriched Exosomes Enhance Neurite Remodeling in the IBZ

Stroke evokes partial damage and disorganization to axon–myelin bundles in the IBZ49. Double staining for Bielschowsky silver and Luxol fast blue was employed to identify axons and myelin in the white matter in the brain, respectively. Axonal density along the IBZ was significantly increased by the exosome treatment compared with the PBS control 28 days after MCAO, and Ex-miR-133b+ further increased the axonal density compared with the Ex-Con (p<0.05, respectively) (Fig. 2, row A).

The phosphorylation of the neurofilament (NF) is considered an important determinant of filament caliber, plasticity, and stability50. Immunocytochemically purified anti-NF-H phosphorylated antibody (clone SMI 31) reacts broadly with thick and thin axons and some dendrites such as basket cell dendrites, but not Purkinje cell

**Table 1.** Relative miR-133b Level in the Various Lentivirus-Infected MSCs and Their Corresponding Exosomes

<table>
<thead>
<tr>
<th>Groups</th>
<th>In Cells (Relative Fold)</th>
<th>In Exosomes (Relative Fold)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Naive MSCs</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>miR-CON+MSCs</td>
<td>1.01±0.21</td>
<td>0.98±0.23</td>
</tr>
<tr>
<td>miR-133b+MSCs</td>
<td>432.65±125.01*</td>
<td>8.83±2.31*</td>
</tr>
<tr>
<td>miR-CON−MSCs</td>
<td>1.02±0.13</td>
<td>1.01±0.34</td>
</tr>
<tr>
<td>miR-133b−MSCs</td>
<td>0.23±0.19†</td>
<td>0.45±0.11†</td>
</tr>
</tbody>
</table>

*p<0.01 compared with the miR-CON+MSCs; †p<0.05 compared with the miR-CON−MSCs.

**Figure 1.** MicroRNA 133b (miR-133b)-enriched exosome treatment enhances neurological outcome improvement. Foot fault test (A) and modified neurologic severity score (mNSS) (B) data show that compared with the phosphate-buffered saline (PBS) treatment, both multipotent mesenchymal stromal cell (MSC) exosome treatment groups exhibited significant improvement of functional recovery (p<0.05 after day 14, respectively). Compared with the MSCs infected with blank vector (Ex-Con), exosomes isolated from miR-133b-overexpressing MSCs (Ex-miR-133b+) significantly increased functional improvement (p<0.05 after day 21, respectively). PBS, middle cerebral artery occlusion (MCAO) rats treated with PBS; CON+, MCAO rats treated with Ex-Con; 133b+, MCAO rats treated with Ex-miR-133b+; EXO, exosomes given. *p<0.05, compared to PBS; #p<0.05 compared to CON+, respectively. Mean±standard error (SE), n=6/group.
Figure 2. MicroRNA 133b (miR-133b)-enriched exosomes further increase neurite remodeling and synaptic plasticity in the ischemic boundary zone (IBZ) compared to control multipotent mesenchymal stromal cell (MSC) exosomes. Bielschowsky silver and Luxol fast blue double staining (A row), SMI 31 immunostaining (B row), and synaptophysin immunostaining (C row) show that based on the increase of neurite remodeling and synaptic plasticity in the IBZ by the MSCs infected with blank vector (Ex-Con) treatment, the exosomes isolated from miR-133b-overexpressing MSC (Ex-miR-133b+) treatment further increased neurite remodeling and synaptic plasticity in the IBZ. PBS, middle cerebral artery occlusion (MCAO) rats treated with phosphate-buffered saline (PBS); CON+, MCAO rats treated with Ex-Con; 133b+, MCAO rats treated with Ex-miR-133b+. *p < 0.05 compared to PBS; #p < 0.05 compared to CON+, respectively. Mean ± standard error (SE), n = 6/group. Scale bars: 100 µm.
and support neural plasticity33,56–58. We therefore measured neuron–glial–vascular communication and other glial cells in the central nervous system (CNS) 28 days after MCAO, and similarly, Ex-miR-133b+ further increased the p-NFH immunoreactivity compared with the Ex-Con (p<0.05, respectively) (Fig. 2, row B).

Immunostaining data also showed that compared with the PBS control, the MSC exosome treatment significantly increased synaptophysin, a presynaptic vesicle protein that identifies synaptic plasticity and synaptogenesis52 in the IBZ 28 days after MCAO, while Ex-miR-133b+ further increased the synaptophysin immunoreactivity compared with the Ex-Con (p<0.05, respectively) (Fig. 2, row C).

**miR-133b-Enriched Exosomes Increase Exosomes in Ischemic Brain**

Cellular release of vesicles containing functional components (e.g., RNAs and proteins) that are taken up and incorporated into the target recipient cell has emerged as a regulatory mechanism for coordination of nervous system function35-55. Exosomes released by astrocytes and other glial cells in the central nervous system (CNS) likely modulate neuron–glial–vascular communication and support neural plasticity35,56–58. We therefore measured the amount of exosomes present in the IBZ. Our data show that the Ex-miR-133b+ treatment significantly increased the presence of exosomes in brain IBZ compared with the Ex-Con treatment (p<0.05) (Fig. 3C) and the PBS treatment.

**miR-133b-Enriched Exosomes Increase the Release of Exosomes From Cultured Primary Astrocytes**

Since astrocytes are the most abundant cells in the nervous system and play an active role in brain recovery59–61, and OGD culture has been used as an in vitro model to mimic the in vivo ischemic condition62–64, we employed OGD-cultured astrocytes to elucidate mechanisms underlying the enhanced therapeutic effects of Ex-miR-133b+ on rats subjected to MCAO. First, we investigated the effects of Ex-miR-133b+ on astrocyte exosomal release. OGD astrocytes were incubated with Ex-Naive, Ex-miR-133b−, and Ex-miR-133b+, as well as exosomes from miR-CON+MSCs and miR-CON−MSCs, and the release of exosomes from astrocytes was quantified. Our data showed that astrocytes subjected to OGD treatment released exosomes (p<0.05) (Fig. 4A). In a complementary experiment, we also investigated the effect of overexpression of miR-133b on MSCs to release exosomes. Infection of MSCs with the blank vector controls did not significantly affect MSCs to release exosomes compared to naive MSCs. However, overexpression or inhibition of miR-133b significantly increased or decreased, respectively, MSC release of exosomes compared to naive MSCs (p<0.05) (Fig. 4B).

**Exosomes Harvested From OGD Astrocytes Treated With Ex-miR-133b+ Increase Neurite Outgrowth**

By employing the primary cortical neuronal neurite outgrowth assay, we tested the effects of OGD astrocyte exosomes on neurite branching and elongation of the neurons. Our data show that exosomes harvested from OGD astrocytes significantly increased the neurite branch number and total neurite length per neuron compared with the non-exosome-treated neurons. Compared to exosomes from astrocytes subjected to OGD only, exosomes harvested from OGD astrocytes pretreated by Ex-Con and Ex-miR-133b+ substantially increased the neurite branch number and total neurite length of
primary cultured cortical neurons, while exosomes from OGD astrocytes treated by Ex-miR-133b+ employed to treat primary cortical neurons had more robust effects on neurite outgrowth compared with the exosomes from OGD astrocytes with Ex-Con (p < 0.05, respectively) (Fig. 5A–D).

miR-133b-Enriched Exosomes Downregulate the Protein Level of RABEPK

MVBs can either be routed to lysosomes for degradation or to exosomes for secretion65. Lysosomal dysfunction increases exosomal release66,67; RABEPK, cooperating with Rab9, participates in the endosome-to-trans-Golgi network (TGN) transport, which is essential for the lysosome formation68. Using Western blot and immunohistochemistry staining of IBZ tissues, we therefore examined whether the exosomal treatment affects RABEPK. Our data revealed that ischemia increased RABEPK protein levels, whereas the Ex-Con treatment significantly reduced MCAO-increased RABEPK proteins and the Ex-miR-133b+ treatment further decreased the protein level (p < 0.05, respectively) (Fig. 6A and B).

DISCUSSION

Exosomes released by transplanted cells mediate the benefits of cell-based therapies for stroke6,8,69,70. It is therefore reasonable to consider replacing cell-based therapy for stroke with cell-free exosome-based treatment, which would minimize potential adverse effects of administering replicating and embolism-forming cells71. We therefore employed exosomes harvested from naive MSCs to treat experimental stroke models and found that the therapeutic effects of exosomes were consistent with the therapeutic effects of direct MSC treatment7. In the current study, we extended this exosome therapy concept to tailor the miRNA content of the exosomes. On the basis of studies indicating that miR-133b promotes neurovascular plasticity, we generated and employed exosomes enriched with miR-133b for the treatment of rats subjected to MCAO. We found that miR-133b-enriched exosomes significantly amplified the therapeutic benefits for stroke compared to control MSC exosomes. Our in vivo and in vitro studies also suggest that astrocytes and possibly other neural parenchymal cells participate in

**Figure 4.** MicroRNA 133b (miR-133b)-enriched exosomes enhance the release of exosomes from primary cultured oxygen- and glucose-deprived (OGD) astrocytes. MSC exosomes significantly increased the exosomes released by OGD astrocytes, and there was no obvious difference on the release from OGD astrocytes, after treatment with harvested from naive MSCs (Ex-Naive) and two control exosomes (A). Compared with the exosome Ex-Naive treatment, exosomes released by OGD astrocytes treated with miR-133b-overexpressing MSCs (Ex-miR-133b+) were significantly increased (A) and significantly decreased (B) by treatment with Ex-miR-133b−, respectively. Correlated with the miR-133b levels in the multipotent mesenchymal stromal cells (MSCs) (Table 1), the number of exosomes released by MSCs has no difference between naive MSCs and the two control MSCs (miR-CON−MSCs and miR-CON+MSCs), but the exosomal release was significantly increased from the miR-133b+MSCs and decreased from the miR-133b−MSCs, compared with that from naive MSCs (B). NivM, naive MSCs; NivA, naive astrocytes; NON, nontreated OGD astrocytes; MSC, OGD astrocytes treated with naive MSC exosomes; CON+, miR-CON+MSC or OGD astrocytes treated with miR-CON+MSC exosomes; 133b+, miR-133b+MSC or OGD astrocytes treated with miR-133b+MSC exosomes; 133b−, miR-133b−MSC or OGD astrocytes treated with miR-CON−MSC exosomes; 133b+, miR-133b+MSC or OGD astrocytes treated with miR-133b+MSC exosomes. *p < 0.05, mean ± SD, n = 3/group.
miR-133b INCREASES BRAIN SECONDARY EXOSOME RELEASE

The enhanced functional recovery for stroke induced by miR-133b-enriched exosomes. miR-133b increased the secondary release of exosomes from astrocytes, which appear to more potently enhance neurite outgrowth and plasticity than naive MSC exosomes. Thus, our study provides the basis for engineering miRNA content of exosomes that concurrently regulate multiple molecular pathways and further promote restorative processes, and that exosomes, whether naive or engineered, induce secondary release of astrocytic exosomes. Here we focused on astrocytes, and we do not exclude the possibility that other parenchymal and inflammatory or vascular cells may also release secondary exosomes in response to primary exosome stimulation.

EVs, including exosomes, are involved in intercellular communication within the nervous system and play major roles in neurological diseases and in nerve trauma, as well as mediate signaling during brain development and function. Exosomes released by brain parenchymal cells modulate neuron–glial–vascular communication and support neural plasticity. By modulating the interactions between glia and neurons, glial-derived exosomes promote neurite outgrowth and neuronal survival under high neuronal activity and/or oxidative stress condition. miR-133b-enriched exosomes increased exosome content within the IBZ. This increase may be attributed to exosomes generated by astrocytes. However, as noted, we cannot exclude the possibility that the preferential increase of exosomes released by other neural parenchymal cells induced by miR-133b-enriched exosomes may also participate in the enhanced neural plasticity and functional recovery, since other brain parenchymal cells (e.g., oligodendrocytes) also secrete exosomes that contain major myelin- and stress-protective proteins, and these exosomes are involved in glia-mediated trophic support to axons.

MVBs are generated by inward budding of the late endosomal membrane, which gives rise to intraluminal

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Figure 5. Exosomes harvested from oxygen- and glucose-deprived (OGD) astrocytes after treatment with miR-133b-overexpressing MSCs (Ex-miR-133b+) increase neurite outgrowth. (A) The morphology of cultured neurons in this assay. Exosomes harvested from OGD astrocytes significantly increased the neurite number and total neurite length per neuron compared with the non-exosome-treated primary cultured cortical neurons. The neurite number and total neurite length substantially increased after the neurons were treated with exosomes harvested from OGD astrocytes that were pretreated by the MSCs infected with blank vector (Ex-Con) and Ex-miR-133b+, and Ex-miR-133b+-pretreated OGD astrocytic exosomes have the enhanced effect compared with the Ex-Con pretreatment (p<0.05, respectively; B–D). Con, non-exosome-treated neurons; OGD, neurons treated with exosomes harvested from OGD astrocytes; Con+, neurons treated with exosomes harvested from OGD astrocytes pretreated with Ex-Con; 133b+, neurons treated with exosomes harvested from OGD astrocytes pretreated with Ex-miR-133b+. *p<0.05, mean±standard deviation (SD), n=3/group. Scale bar: 50 µm.
Figure 6. MicroRNA 133b (miR-133b)-enriched exosomes downregulate the protein level of Rab9 effector protein with kelch motifs (RABEPK). Western blot data show that the MSCs infected with blank vector (Ex-Con) treatment significantly downregulated the RABEPK level increased in the ischemic boundary zone (IBZ) after middle cerebral artery occlusion (MCAO), and miR-133b-overexpressing MSC (Ex-miR-133b+) treatment further reduces the protein level (A, B) \( (p<0.05, \text{ respectively}) \). Double immunostaining shows the RABEPK largely expressed in the astrocytes (E–H). Ex-Con treatment significantly reduced the RABEPK level increase in the astrocytes subjected to oxygen and glucose deprivation (OGD), and Ex-miR-133b+ treatment further reduces the protein level (C, D) \( (p<0.05, \text{ respectively}) \). Sham, tissue from sham surgery rats; PBS, MCAO rats treated with phosphate-buffered saline (PBS); Naive, naive astrocytes; OGD, astrocytes subjected to OGD; CON+, MCAO rats or OGD astrocytes treated with Ex-Con; 133b+, MCAO rats or OGD astrocytes treated with Ex-miR-133b+. \*\( p<0.05 \), mean ± standard deviation (SD), \( n=3\)/group. Scale bar: 50 µm.
vesicles (ILVs) within late endosomes. MVBs can be either routed to lysosomes for degradation or fused with the plasma membrane to discharge their ILVs as exosomes to the extracellular space. Delivery of lysosomal enzymes is essential for the maintenance of proper functioning lysosomes, which include addition of a specific lysosomal tag, mannose 6-phosphate (M6P), to peptides, for example, lysosomal hydrolases. With binding of these tags to M6P receptors in the Golgi apparatus and proper packaging into vesicles destined for the lysosomal system, the hydrolases are released in the acidic environment of endosomes, and the receptor is retrieved to the Golgi by retromer and Rab. RABEPK is required for this endosome-to-TGN transport, so the lack of RABEPK may lead to reduced lysosome formation. Lysosomal dysfunction causes increased exosomal release. Our data suggest that increased RABEPK level in the IBZ may consequently increase the lysosome formation. These data are consistent with prior studies showing that the autophagy and lysosomal degradation pathways are stimulated in hypoxia–ischemia-induced brain injury. Endocytosis and autophagy cooperate and share molecular machinery, such as the Rab GTPase network. The structure of the autophagy pathway, the autophagosome, can fuse with endocytic structures such as MVBs to generate the amphisome, which fuses with the lysosome to degrade the material wrapped inside. Hypoxia induces exosome release as well. Importantly, our data indicated that the therapeutic effects were mediated by exosome transfer of miR-133b from the MSCs to the brain. In the present study, our primary goal was to demonstrate, as a proof of principle, that we can further improve functional recovery by administering tailored exosomes enriched in miR-133b. Further studies, in which miR-133b exosome content is reduced to verify the function of exosomal miR-133b, are warranted. Although volumes of cerebral infarction are not altered by the MSCs and MSC exosome treatment, we cannot exclude the possibility that treatment with exosomes harvested from miR-133b-overexpressing MSCs at 24 h poststroke also evokes some neuroprotective benefits in the ischemic boundary region. CONCLUSIONS IA-administered exosomes harvested from miR-133b-overexpressing MSCs improve neural plasticity and functional recovery after stroke beyond that of naive exosomes. Exosomes enriched with mature miR-133b from miR-133b-overexpressing MSCs further stimulate and increase exosomes release from astrocytes, possibly by downregulating the RABEPK expression. The exosomes derived from OGD astrocytes pretreated with miR-133b-enriched exosomes promote neurite outgrowth and elongation in vitro. We thus propose that the contribution of the secondary astrocytic exosome release may contribute to brain plasticity and subsequent neurological recovery after stroke.

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